



EVALUATION OF *IN VITRO* FREE RADICAL SCAVENGING EFFICACY OF DIFFERENT ORGANIC EXTRACTS OF *MORINDA TINCTORIA* LEAVES

K.P. SREENA¹, A. POONGOTHAI¹, S.V. SOUNDARIYA, ¹ G. SRIREKHA, ¹ R. SANTHI ² AND S. ANNAPOORANI³

¹Research Scholars, ²Assistant Professor in Biotechnology, ³Professor in Biochemistry, ^{1,3}Avinashilingam Deemed University for Women, Coimbatore - 43. ²Dr.GRD College of Science, Coimbatore - 14, Tamilnadu, India. Email:santhiakash@yahoo.com

Received: 11 March 2011, Revised and Accepted: 10 April 2011

ABSTRACT

In vitro free radical scavenging efficacy of methanolic, ethanolic, and ethylacetate extracts of *Morinda tinctoria* leaves were carried out. *In vitro* free radical scavenging efficacies of these extracts were assessed by studying its ability to scavenge DPPH, NO, H₂O₂ and Hydroxyl radicals. The methanolic extract of *Morinda tinctoria* leaves showed IC₅₀ with minimum concentration and more effective in scavenging DPPH, NO, H₂O₂ and Hydroxyl radicals followed by ethanol and ethyl acetate extracts respectively. Thus, the methanolic extract of *Morinda tinctoria* leaves can be recommended as a potent antioxidant in patients suffering from various oxidative degenerative diseases such as cancer, diabetes, arthritis, and cardiovascular diseases. This free radical scavenging activity might be due to the active components present in the methanolic extract of *Morinda tinctoria* leaves.

Keywords: *Morinda tinctoria*, Free radical scavenging activity.

INTRODUCTION

It is increasingly being realized that many of today's diseases are due to the "oxidative stress" that results from an imbalance between formation and neutralization of prooxidants. Oxidative stress is initiated by free radicals, which seek stability through electron pairing with biological macromolecules such as proteins, lipids and DNA in healthy human cells and cause protein and DNA damage along with lipid peroxidation. These changes contribute to cancer, atherosclerosis, cardiovascular diseases, ageing and inflammatory diseases¹. The free radicals are known to be scavenged by synthetic antioxidants, but due to their adverse side effects leading to carcinogenicity; search for effective and natural antioxidants has become crucial². In recent times, focus on plant research has increased all over the world and a large body of evidences has collected to show immense potential of medicinal plants used in various traditional systems³. Plants are endowed with free radical scavenging molecules, such as vitamins, terpenoids, phenolic acids, lignins, stilbenes, tannins, flavonoids, quinones, coumarins, alkaloids, amines, betalains and other metabolites, which are rich in antioxidant activities⁴. A number of plants and plant isolates have been reported to protect free radical induced damage in various experimental models⁵.

Morinda tinctoria (family Rubiaceae) is commercially known as *Nunaa* and is indigenous to tropical countries⁶. In the traditional system of medicine, leaves and roots of *Morinda tinctoria* are used as astringent, deobstruent, emmenagogue and to relieve pain in the gout⁷. There is greater demand for fruit extract of *morinda* species in treatment for different illness such as arthritis, cancer, gastric ulcer and other heart diseases⁸. Anti convulsant, analgesic and anti inflammatory activities of *Morinda tinctoria* has been reported^{9,10}. Hence the present study has been made to investigate the *in vitro* free radical scavenging efficacy of different organic extracts of *Morinda tinctoria* leaves.

MATERIALS AND METHODS

Collection of plant material

Fresh leaves of *Morinda tinctoria* were collected from the outskirts of Coimbatore district, Tamilnadu. The collected leaves were washed thoroughly in tap water, shade dried and finely powdered.

Preparation of methanolic, ethanolic and ethyl acetate extracts of *Morinda Tinctoria* leaves

10g of powder of *Morinda tinctoria* was filled in the thimble and extracted with 150ml of methanol, ethanol and ethyl acetate using a soxhlet extractor for 24 hours. The extracts were then distilled and

evaporated to dryness. The concentrated extracts were then accurately weighed and stored in small vials at -20°C, for further studies.

Assessment of *in vitro* free radical scavenging efficacy of different organic extracts of *Morinda Tinctoria* leaves DPPH radical scavenging assay

This was assayed as described by Elizabeth and Rao (1990)¹¹. The reaction mixture contained Methanol-50 ml. DPPH (Diphenyl-2-picryl hydrazyl radical)-1mM 3 ml of 1mM DPPH in methanol was added to 100µl of plant extract with concentrations ranging from 10µg to 100µg. DPPH solution with methanol was used as a positive control and methanol alone acted as a blank. When DPPH reacts with antioxidant in the sample, it was reduced and the color changed from deep violet to light yellow. This was measured at 518 nm. The percentage scavenging activity was calculated by the following formula.

$$\text{Scavenging activity (\%)} = \frac{A518(\text{control}) - A518(\text{sample})}{A518(\text{control})} \times 100$$

Nitric oxide radical scavenging activity

Nitric oxide generated from sodium nitroprusside in aqueous solution at physiological pH interacts with oxygen to produce nitrite ions, which were measured by the method of Garrat (1964)¹². The reaction mixture (3ml) containing 2 ml of sodium nitroprusside (10mM), 0.5 ml of phosphate buffer saline (1M) were incubated at 25°C for 150 mins. After incubation, 0.5 ml of the reaction mixture containing nitrite was pipetted and mixed with 1 ml of sulphanic acid reagent (0.33%) and allowed to stand for 5 min for completing diazotization. Then 1 ml of naphthylethylene diamine dihydrochloride (1% NEDA) was added, mixed and allowed to stand for 30 mins. Sodium nitroprusside in aqueous solution at physiological pH spontaneously generates nitric oxide, which interacts with oxygen to produce nitrite ions which can be estimated by the use of Griess illosvery reaction at 540 nm.

Hydrogen peroxide scavenging assay

Hydrogen peroxide assayed as described by (Ruch *et al.*, 1989)¹³ proposed an assay for the determination of antioxidant activity of compounds by their ability to scavenge the oxidant hydrogen peroxide. The reaction mixture contained Phosphate buffer (pH-7.4) hydrogen peroxide in phosphate buffer (40mM). A solution of hydrogen peroxide (40mM) was prepared in phosphate buffer. Plant extracts at the concentration of 10mg/10µl was added to a hydrogen peroxide solution (0.6ml,40mM). The total volume was made up to 3ml. The absorbance of the reaction mixture was recorded at

230nm. The blank solution contained phosphate buffer without hydrogen peroxide. The percentage of hydrogen peroxide scavenged by the plant extract was calculated as follows:

$$\text{Percentage of scavenged H}_2\text{O}_2 = \frac{A_0 - A_1 \times 100}{A_0}$$

A₀- Absorbance of control

A₁- Absorbance in the presence of plant extract

Hydroxyl radical scavenging activity

Hydroxyl radical assayed as described by Elizabeth and Rao (1990)¹⁴. The assay is based on quantification of degradation product of 2-deoxy ribose by condensation with TBA. Hydroxyl radical was generated by the Fe³⁺-Ascorbate - EDTA - H₂O₂ system (Fenton reaction). The reaction mixture contained 0.1 ml deoxyribose (2.8mM), 0.1 ml EDTA (0.1 mM), 0.1 ml H₂O₂ (1mM), 0.1 ml Ascorbate (0.1mM), 0.1 ml KH₂PO₄-KOH buffer, pH 7.4 (20mM) and various concentrations of plant extract in a final volume of 1 ml. The reaction mixture was incubated for 1 hour at 37°C. Deoxyribose degradation was measured as TBARS and the percentage inhibition was calculated.

RESULTS

In vitro free radical scavenging efficacy of various organic extracts of *Morinda tinctoria* leaves were assayed by its ability to scavenge DPPH, NO, H₂O₂ and OH radicals. Fig 1 illustrates the dose response curve of DPPH radical scavenging activity of methanolic, ethanolic and ethyl acetate extracts of *Morinda tinctoria* leaves. The IC₅₀ values of methanolic, ethanolic and ethyl acetate extracts were found to be 47µg, 51µg and 58µg respectively. It was observed that methanolic extract of *Morinda tinctoria* leaves had higher activity for scavenging DPPH than that of ethanolic and ethyl acetate extracts.

Methanolic extract of *Morinda tinctoria* leaves showed the highest NO scavenging activity when compared to ethanolic and ethyl acetate extracts in a moderate dose dependent manner with an IC₅₀ value of 45 µg as shown in the Fig 2. The IC₅₀ values of ethanolic and ethyl acetate extracts were found to be 59µg and 57µg respectively.

Scavenging of H₂O₂ and its percentage inhibition of methanolic, ethanolic and ethyl acetate extracts of *Morinda tinctoria* leaves showed the IC₅₀ values of 32µg, 40 µg and 45 µg respectively. The results are shown in the Fig 3. Here also the methanolic extract of *Morinda tinctoria* leaves showed IC₅₀ with minimum concentration followed by ethanolic and ethyl acetate extracts.

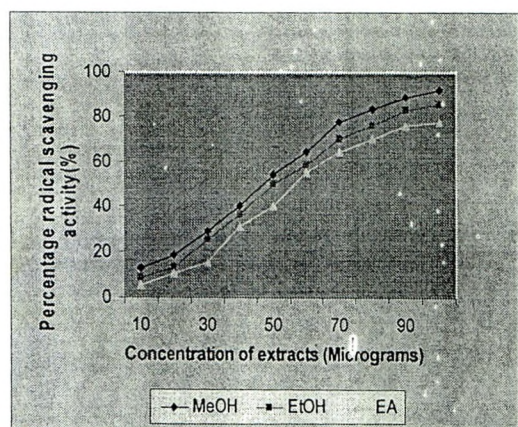


Fig. 1: Dpph radical scavenging efficacy of various organic extracts of *Morinda tinctoria* leaves

The OH radical scavenging assay shows the ability of methanolic, ethanolic and ethyl acetate extracts to inhibit OH radical mediated deoxyribose degradation in an Fe³⁺-EDTA- Ascorbic acid - H₂O₂ reaction mixture. The results are shown in the Fig 4. The IC₅₀ values

of the methanolic, ethanolic and ethyl acetate extracts were 27 µg, 55 µg and 62 µg respectively. Here also the methanolic extracts of *Morinda tinctoria* leaves showed IC₅₀ with minimum concentration followed by ethanol and ethyl acetate extracts.

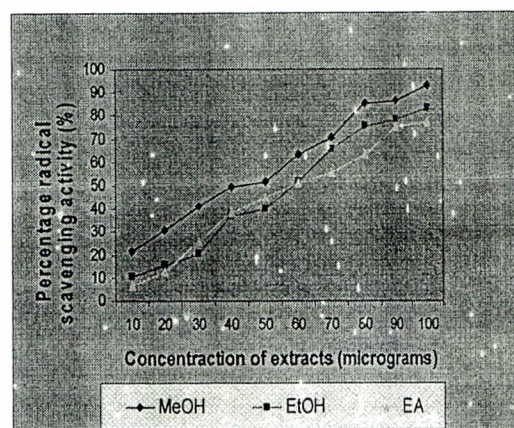


Fig. 2: Nitric oxide radical scavenging efficacy of various organic extracts of *Morinda tinctoria* leaves

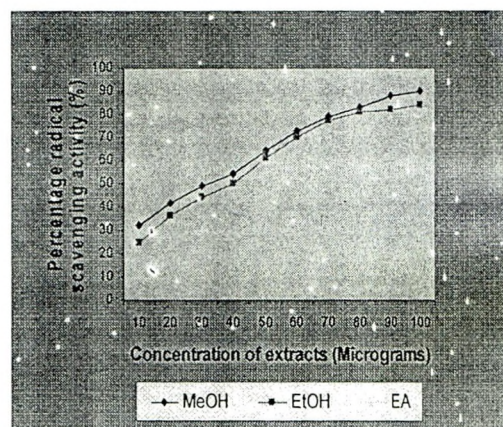


Fig. 3: Hydrogen peroxide radical scavenging efficacy of various organic extracts of *Morinda tinctoria* leaves

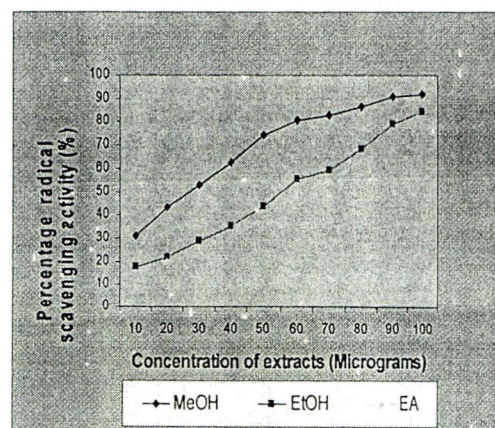


Fig. 4: Hydroxyl radical scavenging efficacy of various organic extracts of *Morinda tinctoria* leaves

DISCUSSION

DPPH radical scavenging is considered a good in vitro model widely used to assess antioxidant efficacy within a very short time. In its radical form, DPPH has disappears on reduction by an antioxidant compound or a radical species to become a stable diamagnetic molecule resulting the colour changes from purple to yellow, which could be taken as an indication of the hydrogen donating ability of the tested sample^{15, 16}. DPPH radical scavenging ability of the methanolic extract of *Morinda tinctoria* leaves were significantly higher than that of ethanolic and ethyl acetate extracts.

It is well known that nitric oxide has an important role in various inflammatory processes. Sustained levels of production of this radical are directly toxic to tissues and contribute to the vascular collapse associated with septic shock, whereas chronic expression of nitric oxide radical is associated with various carcinomas and inflammatory conditions including juvenile diabetes, multiple sclerosis, arthritis and ulcerative colitis¹⁷. The toxicity of NO increases greatly when it reacts with super oxide radical, forming the highly reactive peroxy nitrite anion (ONOO⁻)¹⁸. The nitric oxide generated from sodium nitro prusside reacts with oxygen to form nitrite. The methanolic extract of *Morinda tinctoria* leaves inhibits the nitrite formation by directly competing with oxygen in the reaction with NO. This study proved that the methanolic extract has more potent NO scavenging activity when compared to ethanolic and ethyl acetate extracts.

H₂O₂ is highly important because of its ability to penetrate biological membranes. H₂O₂ itself is not very reactive, but it can sometimes be toxic to cell because it may give rise to hydroxyl radical in the cells¹⁹. The results showed that the methanolic extract of *Morinda tinctoria* leaves had an effective H₂O₂ scavenging activity.

Hydroxyl radicals are most reactive species, initiating the peroxidation of the cell membrane²⁰. The lipid radical, thus generate would initiate chain reaction in the presence of oxygen, giving rise to lipid peroxide, which break down to aldehydes such as malondialdehyde. The IC₅₀ value indicates that the methanolic extract of *Morinda tinctoria* leaves are better hydroxyl radical scavenger than that of ethanolic and ethyl acetate extracts.

CONCLUSION

On the basis of the results obtained in the present study, it concludes free radical scavenging efficacy of methanolic extract of *Morinda tinctoria* leaves and thus gives scientific basics for its traditional uses as potent antioxidant to the individuals under oxidative stress such as inflammation, aging, mutagenicity and carcinogenicity. In future we look forward to check the potency of methanolic extract of *Morinda tinctoria* leaves by means of *in vivo* antioxidant studies and also to isolate, identify and characterize the active constituents responsible for these effects.

REFERENCES

1. Bibhabasu H, Santanu B, Nripendranath M, Antioxidant and free radical scavenging activity of *Spondias pinnata*, BMC Complementary and Alternative Medicine, Vol 8, p.63,2008.
2. Akiri SVCR, Sareddy GR, Phanithi PB, Attipalli RR, The antioxidant and antiproliferative activities of methanolic extracts from Njavara rice bran, BMC Complementary and Alternative Medicine, Vol 10, p.4, 2010.
3. Ashish JM, Khadabadi SS, Deokate UA, Farooqui IA, Deore SL, Gangwani MR, *Argyria speciosa* Linn.f. Phytochemistry, pharmacognosy and pharmacological studies, Journal of pharmacognosy and phytotherapy, Vol 2, p.34-42, 2010.
4. Aiyegoro OA, Okoh AI, Preliminary phytochemical screening and *in vitro* antioxidant activities of the aqueous extract of *Helichrysum longifolium* DC, BMC Complementary and Alternative Medicine, Vol 10, p.21, 2010.
5. Himakar RK, Tharanath V, Nagi RKB, Sharma PVGK, Reddy OVS, Studies on hepatoprotective effect of hexane extract of *Dillenia indica* against CCl₄ induced toxicity and its safety evaluation in wistar rats, Research journal of Pharmaceutical, Biological and Chemical Sciences, Vol. 1, p.441-450, 2010.
6. Muralidharan P, Sivaraman D, Antihyperglycemic and antidiabetic effects of *Morinda tinctoria* Roxb using streptozotocin - induced diabetic rats, Asian Biomedicine, Vol.3, p.433- 437, 2009.
7. Sivaraman D, Muralidharan P, Evaluation of antimicrobial and anti inflammatory activities of *Morinda tinctoria* Roxb, Asian J.Exp. Biol.Sci., Vol. 1, p.8-13, 2010.
8. Mathivanan N, Surendiran G, Srinivasan K, Malarvizhi K, Morinda pubescens J.E. Smith (*Morinda tinctoria* Roxb.) Fruit extract accelerates wound healing in rats, J. Med. Food, Vol. 9, p.591-593, 2006.
9. Kumaresan PT, Saravanan A, Anticonvulsant activity of *Morinda tinctoria* Roxb, Afr. J. Pharm. Pharmacol., Vol. 3, p.63-65, 2009.
10. Jeyabalan S, Palayan M, Analgesic and anti-inflammatory activity of leaves of *Morinda tinctoria* Roxb, Int.J. Pharm Res., Vol. 1, p.74-80, 2009.
11. Elizabeth K, Rao MWA, Oxygen radical scavenging activity of Curcumin, Int. J. Pharmaceu., Vol. 58, p.237-240, 1990.
12. Garret DC, The quantitative analysis of drugs, Champman and Hall, Japan, Vol. 3, p.456-458, 1964.
13. Ruch RJ, Cheng SJ, Klaunig JE, Prevention of cytotoxicity and inhibition of intracellular communication by antioxidant catechins isolated from Chinese green tea, Carcinogenesis, Vol. 10, p.1003-1008, 1989.
14. Elizabeth K, Rao MWA, Oxygen radical scavenging activity of Curcumin, Int. J. Pharmaceu., Vol. 58, p.237-240, 1990.
15. Marxen K, Vanselow KH, Lippemeier S, Hintze R, Ruser A, Hansen UP, Determination of DPPH radical oxidation caused by methanolic extracts of some micro algal species by linear regression analysis of spectrophotometric measurements, Sensors, Vol. 7, p.2080-2095, 2007.
16. Lee YR, Woo KS, Kim KJ, Son JR, Jeong HS, Antioxidant activities of ethanol extracts from germinated specially rough rice, Food Sci Biotechnol., Vol. 16, p.765-770, 2007.
17. Tylor BS, Kion YM, Wang QI, Sharpio RA, Billiar TR, Geller DA, Nitric oxide down regulates hepatocyte- inducible nitric oxide synthase gene expression, Arch Surg, Vol 132, p.1177-1183, 1997.
18. Huie RE, Padmaja S, The reaction of nitric oxide with superoxide, Free Radic Res Commun., Vol. 18, p.195-199, 1993.
19. Arulmozhi S, Papiya MM, Purnima A, Sathiya N, *In Vitro* Antioxidant and Free Radical Scavenging Activity of *Alstonia scholaris* Linn. R.Br, Iranian Journal of Pharmacology and Therapeutics, Vol. 6, p.191-196, 2008.
20. Halliwell B, Gutteridge JMC, Arnoma OL, The Deoxyribose method: A simple test tube assay for the determination of rate constant for reaction of hydroxyl radical, Anal Biochem., Vol. 165, p.215, 1987.